

Studies on Diversity of Cellulolytic fungi in Gorakhpur, with special reference to cellulase activity of most Frequent fungi

Shailendra Yadav, Supriya Pandey and *Neeraj Srivastava

Applied Mycology Lab, Department of Botany,
St. Andrew's Post-Graduate College,
GORAKHPUR – 273001 (UP), INDIA
*Corresponding Author
E-mail : mycology1999@gmail.com

Received : 11.01.2025; **Revised :** 25.01.2025; **Accepted :** 09.03.2025

How to cite : Yadav S, Pandey S, Srivastava N. Studies on Diversity of Cellulolytic fungi in Gorakhpur, with special reference to cellulase activity of most Frequent fungi. *Flora and Fauna* 2025. 31(1) : 113-120.

ABSTRACT

Microorganisms, especially fungi and bacteria cause biodeterioration of our cultural heritage including paper, cloths, wood and leather. Cellulolytic Fungi cause decomposition of papers and cloths present as waste material. In the present investigation, fungi invading books of Rajkiya Jila Pustkalaya (Govt. District Library), Gorakhpur and Central Library of St. Andrew's College, Gorakhpur have been collected, cultured and identified. Isolations from a total number of eight samples and four isolates from each sample have been done. Fifteen species of these cellulolytic fungi belonging to seven genera have been reported. The two most abundant species found are *Aspergillus niger* and *A. flavus*, which have been isolated from all the 4 isolates of 8 samples. These two species have been selected for detailed studies of their cellulolytic activities. Enzymatic Index (EI) of these two species have been calculated using CMC Agar Medium and Gram Iodine Solution as indicator. EI of *Aspergillus niger* is reported to be 2.50 and of *A. flavus* as 1.3125.

Figures : 07

References : 31

Table : 01

KEY WORDS : *Aspergillus*, Cellulase activity, Cellulolytic fungi, Enzymatic index (EI), Government Library, St. Andrew's College Library.

Introduction

The humid tropical and sub-tropical countries are facing problem of biodegradation of important commodities as waste material. Our cultural heritages are made up mainly of paper, cloth, wood and leather. The hot and humid climate supports growth of microbes including cellulolytic fungi. Movable and immovable, both objects of our cultural heritage are destroyed by these microbes, including fungi and bacteria. Fungi have been reported as the most active microbes and cause maximum damage to these objects¹⁶. Fungi degrading objects containing cellulose, such as papers and cloths, are known as Cellulolytic Fungi. They produce cellulase enzyme and cause biodeterioration of objects in our

cultural heritage, libraries etc.^{18, 30}. High relative humidity and moderate temperature and light intensity of July to October months support abundant growth and decomposition action of these fungi. Cellulolytic fungi like *Alternaria*, *Aspergillus*, *Chaetomium*, *Cladosporium*, *Penicillium*, and *Trichoderma* etc. have been reported to cause decomposition of antique documents^{10, 14, 20, 21, 29}.

Fungal biodeterioration of properties made of papers and archival materials is very high in India². Decomposition of papers have been reported to be caused by a large number of cellulolytic fungi⁵. Infestation of these objects containing cellulose has been reported to be caused by Xerophilic fungi even under very low

ACKNOWLEDGEMENTS : The authors are thankful to National Fungal Culture Collection of India (NFCCI), Agharkar Research Institute, Pune (Maharashtra) for depositing and providing Accession Numbers to pure cultures of *Aspergillus niger* and *A. flavus*; to Indian Type Culture Collection (ITCC), Indian Agricultural Research Institute (IARI), New Delhi for supply of pure culture and to the Principal, St. Andrew's College, Gorakhpur for infrastructural facilities.

TABLE- 1 : Frequencies of Cellulolytic Fungi isolated from 8 Samples of Papers

S. No.	Cellulolytic Fungi	* Frequencies			
		Isolation 1	Isolation 2	Isolation 3	Isolation 4
1.	<i>Alternaria alternata</i>	√	√√	x	x
2.	<i>Aspergillus nidulans</i>	x	√√	√√	x
3.	<i>A. flavus</i>	√√√√	√√√	√√√	√√√
4.	<i>A. fumigatus</i>	√	√√	x	√√
5.	<i>A. terreus</i>	√	√√	x	x
6.	<i>A. ochraceous</i>	√	√	x	x
7.	<i>A. niger</i>	√√√	√√√	√√√	√√√
8.	<i>A. versicolor</i>	x	x	x	√
9.	<i>Chaetomium globosum</i>	√√	√√√	√√	√√√
10.	<i>Cladosporium cladosporioides</i>	√√	√	x	√
11.	<i>C. herbarum</i>	√	√√	√	x
12.	<i>Curvularia lunata</i>	√√	√√√	√√	√√√
13.	<i>Helminthosporium sp.</i>	x	√√	x	x
14.	<i>Penicillium chrysogenum</i>	√√	x	√	√
15.	<i>P. citrinum</i>	√	x	√	√

* x = Absent (No fungal Species), √ = Mild Fungal Growth, √√ = Moderate Fungal Growth, √√√ = Rich Fungal Growth



Fig. 1 : Collection of Cellulolytic Fungi from infested papers

humid conditions. In earlier reports also, rich fungal diversity has been found in library books in Gorakhpur^{27, 28}.

These cellulolytic fungi produce cellulase enzymes to cause biodeterioration of objects like papers and textiles containing cellulose. Nine fungal species have been reported to cause biodegradation of Nineteenth -Century Islamic and Koranic books, with dominance of *Aspergillus niger*, *A. oryzae* and *Hypocrea lixii*. These fungal species showed their ability to degrade Carboxy Methyl Cellulose (CMC)¹³. Workers^{19,21} described four species of these fungi, named *Aspergillus versicolor*, *A. nidulans*, *A. terreus* and *Chaetomium globosum* causing damage of old documents. The cellulase producing ability of these fungi has been exploited for treatment of agro-industrial residues^{4, 6} and

production of cellulase enzymes⁹. Similarly, exploitation of micro-organisms/their enzymes for degradation of lignocellulosic wastes is the need of time²⁰. Protection and preservation of ancient documents from biodegradation is a big challenge²⁶. However, isolation of these fungi causing biodeterioration and their clear identity is the first step in this direction. Only then their cellulolytic activities can be determined using the parameter of Enzymatic Index (EI), and proper and effective control measures can be proposed.

Keeping these facts in view, the present work was undertaken and samples of papers infested with fungi have been investigated and fungal samples have been collected. Various species of cellulolytic fungi have been identified and cellulolytic activities of the most frequent species have been calculated by calculating their Enzymatic Index (EI).

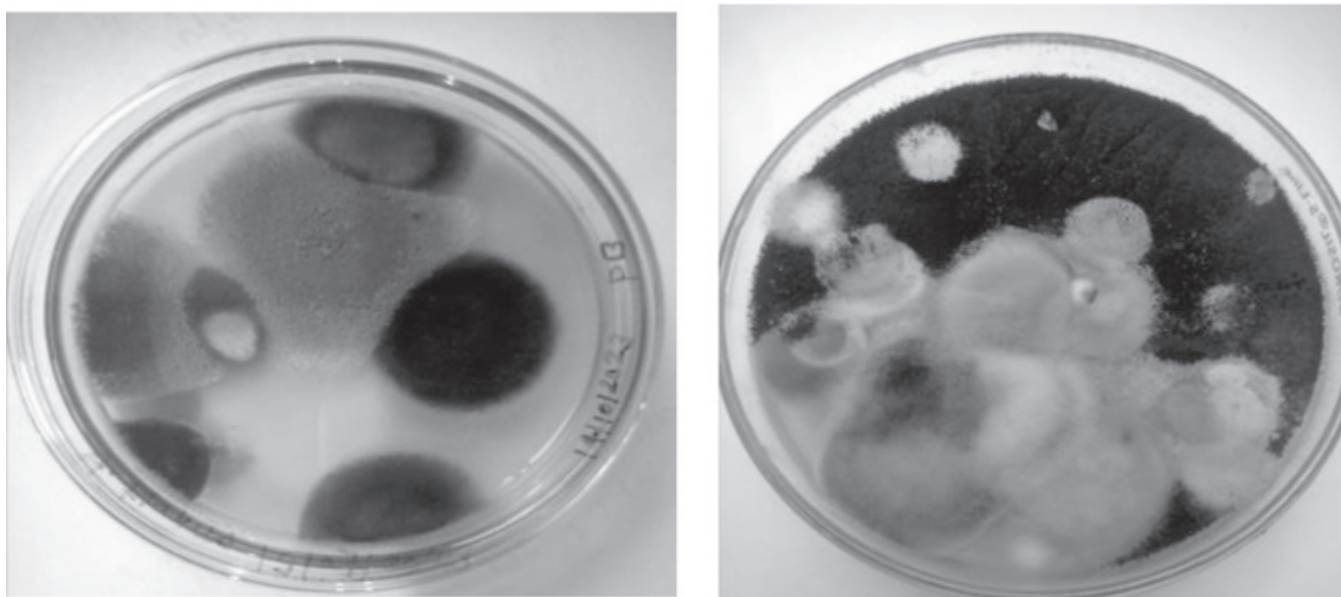


Fig. 2 : Mixed culture obtained from collected samples

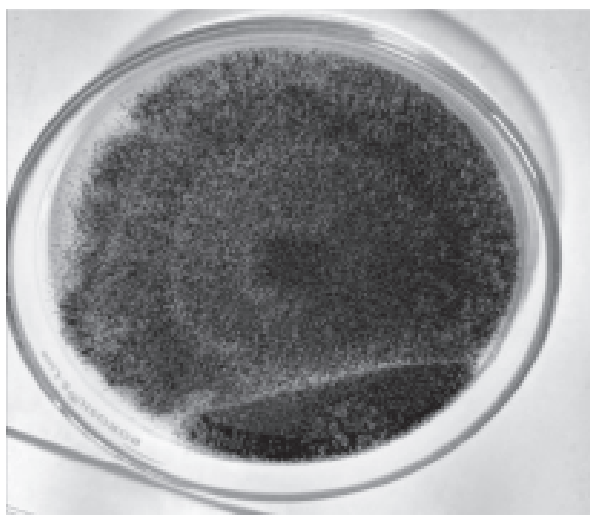


Fig. 3 (A) : *Aspergillus niger*
(Accession No. NFCCI 5688)

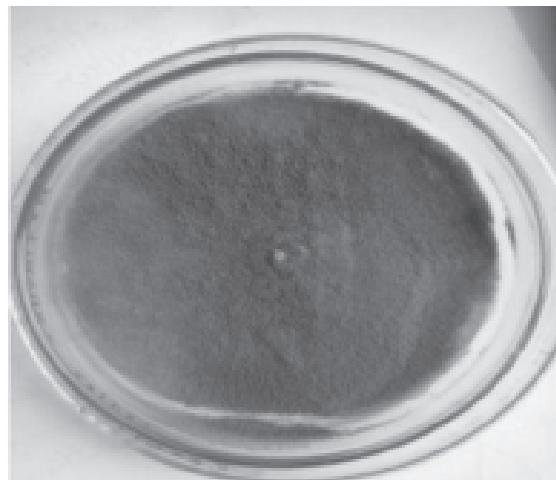


Fig. 3 (B) : *Aspergillus flavus*
(Accession No. NFCCI 5710)
(Pure Culture/ITCC No. 8691)

Pure cultures obtained from mixed culture / ITCC, IARI, New Delhi

Materials and Methods

(1). Collection and Identification of Cellulolytic Fungi

Eight samples (with four isolations from each sample) of cellulolytic fungi were collected from deteriorated Glazed and Unglazed papers in the areas under investigations *i.e.*, Rajkiya Jila Pustkalay (Government District Library), Gorakhpur and Central Library of St. Andrew's Post-Graduate College, Gorakhpur, using sterilized inoculation needle and cotton swab. These fungi were mounted in Lactophenol-Cotton Blue and were examined by direct observation under compound microscope. *In - vitro* culture of these fungi was done on PDA medium. The mixed culture so obtained was purified by streaking and fungi were identified by observing under light microscope and using

various taxonomic criteria. The literatures were used for identification^{3,4,8,11,15,17,23}.

(2). Screening of Cellulolytic Activity

Of all the isolated cellulolytic fungi, the two most frequently occurring fungi were *Aspergillus niger* and *A. flavus*. Accession Numbers of these two fungal species were obtained from National Fungal Culture Collection of India(NFCCI), Pune, Maharashtra (*A. niger* : NFCCI5688; *A. flavus* : NFCCI 5710). The pure culture of *A. flavus* was also obtained from Indian Type Culture Collection (ITCC), IARI, New Delhi (ITCC No. 8691).

These two *Aspergillus* species producing Cellulase Enzyme were selected for screening of their cellulolytic activities, which was done using 1% Carboxy Methyl Cellulose (CMC) Agar Medium. This Carboxy

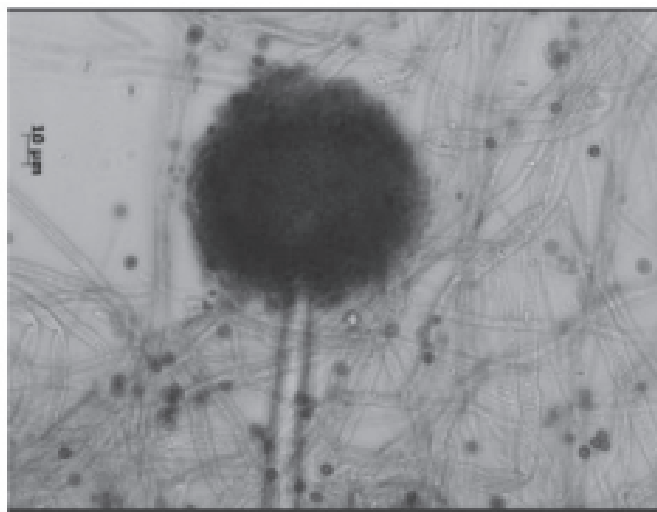
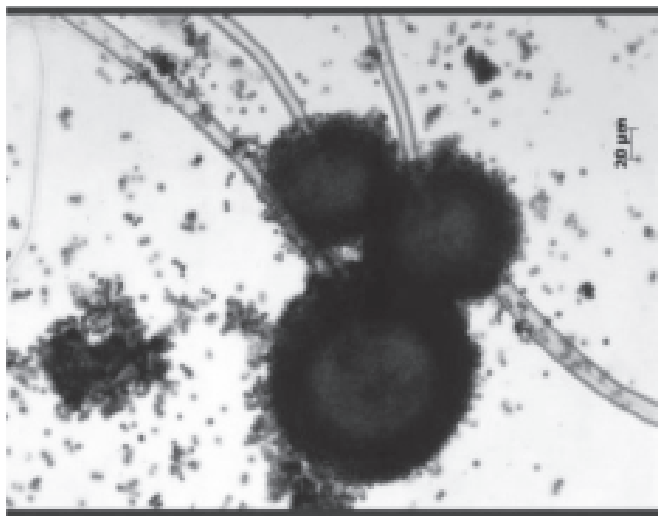
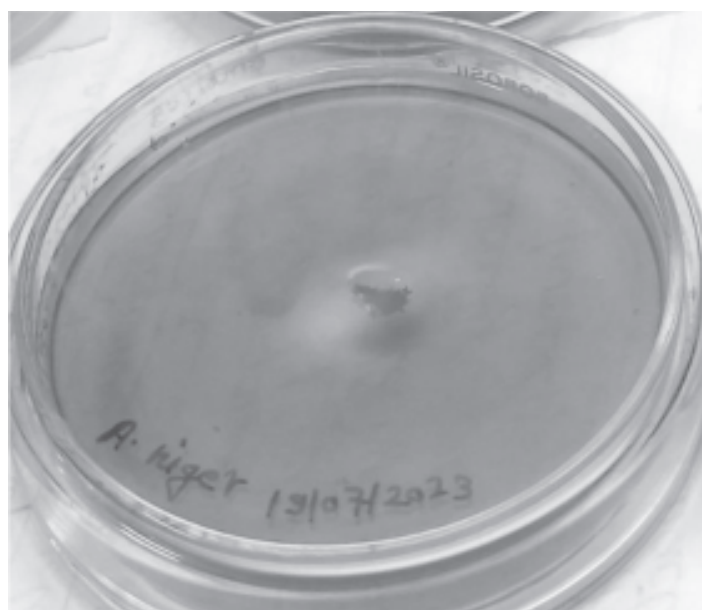
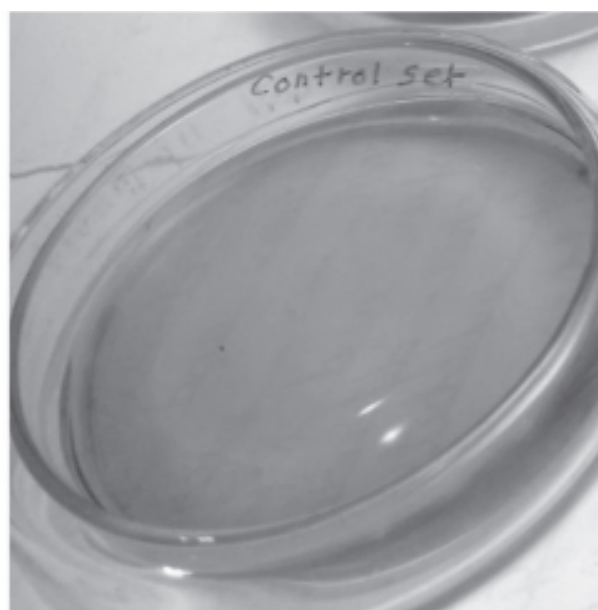


Fig. 4 : *Aspergillus niger* (Mycelium, Conidiophore and Conidia)



Treatment Set (Inoculated)



Control Set (Uninoculated)

Fig. 5 : Culture of *Aspergillus niger* on 1% CMC Agar Medium (Flooded with Gram's Iodine Solution)

Enzymatic Index (EI) of *Aspergillus niger*

Hydrolysis Zone = 25 mm. Diameter

Colony = 10 mm. Diameter

$$\text{Enzymatic Index of } \textit{Aspergillus niger} = 25/10 = 2.50$$

Methyl Cellulose was added as source of Carbon and indicator used was Gram's Iodine Solution¹⁰.

The theory behind this qualitative determination of cellulolytic activities is that, the Gram's Iodine reacts

with both, the cellulose and its degraded components. The integral and unhydrolyzed Cellulose holds the colour of Gram's Iodine dye, whereas the cellulose hydrolyzed into its components by fungal enzymes shows clear zone or pale hallow zone. This clear/pale halo zone was

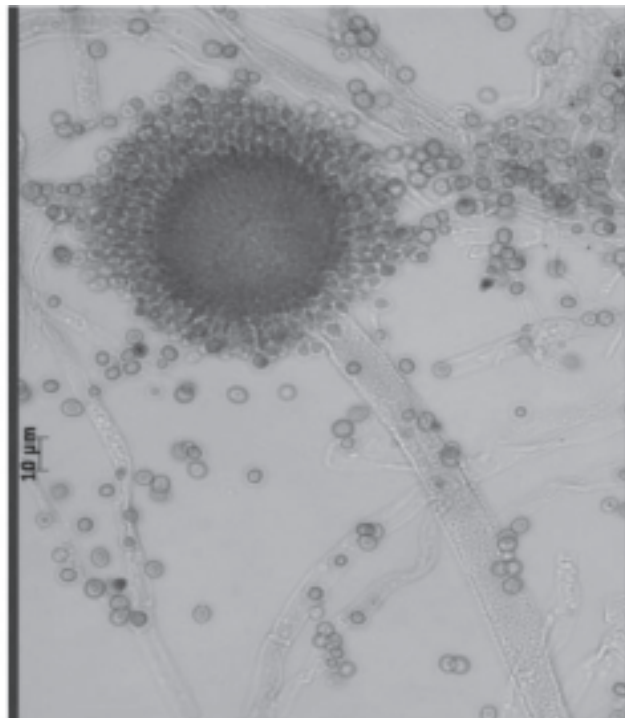
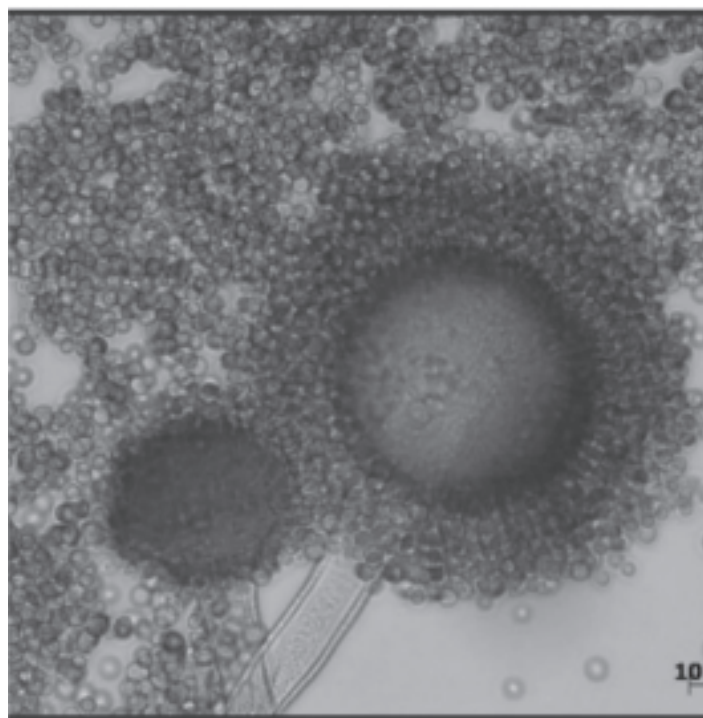
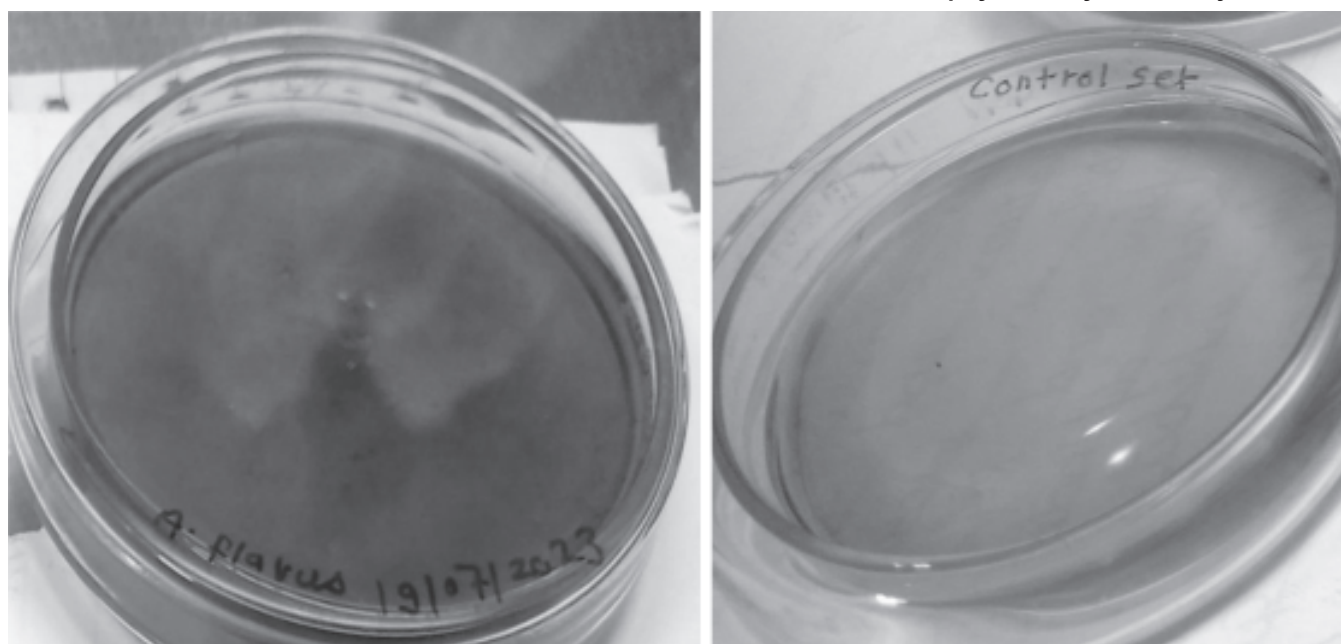


Fig. 6 : *Aspergillus flavus* (Mycelium, Conidiophore and Conidia)



Treatment Set (Inoculated)

Control Set (Uninoculated)

Fig. 7 : Culture of *Aspergillus flavus* on 1% CMC Agar Medium (Flooded with Gram's Iodine Solution)

Enzymatic Index (EI) of *Aspergillus flavus*

Hydrolysis Zone = 21 mm. Diameter Colony = 16 mm. Diameter

Enzymatic Index of *Aspergillus flavus* = $21/16 = 1.3125$

measured and Enzymatic Index (EI) was calculated using the following formula:

$$EI = \frac{\text{Diameter of Hydrolysis Zone}}{\text{Diameter of Colony}}$$

The fungal discs of 4 mm diameter were cultured on 1% CMC Agar Medium containing Streptomycin and were incubated for 7 days at $30 \pm 2^\circ\text{C}$ temperature. After 7 days incubation period, these cultures were treated with Gram's Iodine indicator. Before observation, these cultures were washed with water. The clear zone around the fungal growth was measured and photographs were taken for reporting.

The experiments were repeated twice with one fungal culture used as control set¹⁰.

Observations

Fifteen species of cellulolytic fungi were isolated from infested glazed and unglazed papers (Table – 1).

Result and Discussion

Fifteen species of cellulolytic fungi have been isolated from 8 samples of deteriorated papers (Table -1). *Aspergillus niger* and *Aspergillus flavus* have been isolated from all the 8 samples and 4 isolates, and therefore, have been reported as the most frequently occurring species. *Chaetomium globosum* and *Curvularia lunata*, show moderate growth and others show mild to moderate growth.

Enzymatic Index (EI) of *Aspergillus niger* is 2.50 and of *Aspergillus flavus* is 1.3125.

These cellulolytic fungi causing biodegradation of papers mostly belong to Fungi Imperfecti of Division – Deuteromycotina. Growth of these fungi is supported by high relative humidity and moderate temperature and light intensity. Also, these fungi show great adaptability under changing environment and can survive under unfavorable environmental conditions.

References

1. Abadias M, Usall J, Anguera M, Solsona C, Viñas I. Microbiological quality of fresh, minimally-processed fruit and vegetables, and sprouts from retail establishments, *International Journal of Food Microbiology*. 2008; **123** : 121–129.
2. Agrawal OP. An overview of problems of biodeterioration of cultural property in Asia. In : Biodeterioration of Cultural Property 3. C. Aranyanak and C. Singhasiri. Eds. *Proceedings of the 3rd International Conference of*

Biodeterioration of Cultural Property, Bangkok. 1995:14-34.

3. Ainsworth GC, Bisby GR. Dictionary of Fungi. Common wealth Mycological Institute, Surrey, U.K., 1971.
4. Ames LM. A Monograph of the Chaetomiaceae. No. 2. The United Army Research and Development Series. 1961.
5. Anwar Z, Gulfaz M, Irshad M. Agro-industrial lignocellulosic biomass a key to unlock the future bio-energy: A brief review. *J. Radiat. Res. Appl. Sci.* 2014; **7**: 163–173.
6. Aranyanak C. Microscopical Study of Fungal Growth on Paper and Textile. In : *Biodeterioration of Cultural Property 3*. C. Aranyanak and A. Singhasiri. Eds. *Proceedings of the 3rd International Conference of Biodeterioration of Cultural Property*, Bangkok. 1995; 83-102.
7. Berlemont R. Distribution and diversity of enzymes for polysaccharide degradation in fungi. *Sci. Rep.* 2017; **7**: 222.
8. Booth C. *Fusarium*: Laboratory Guide to the Identification of the Major Species. Common wealth Mycological Institute, Surrey, U.K. 1977.
9. Chiranjeevi T. *et al.* Optimization of holocellulolytic enzymes production by *Cladosporium cladosporioides* using Taguchi-L'16 orthogonal array. *J. Biobased Mater. Bioenergy.* 2012; **6** : 1–10.
10. Coronado-Ruiz C, Avendaño R, Escudero-Leyva E, Conejo-Barboza G, Chaverri P, Chavarría M. Two new cellulolytic fungal species isolated from a 19th century art collection. *Sci. Rep.* 2018; **8** : 7492: 1-9.
11. Ellis MB. Dematiaceous Hyphomycetes. Common wealth Mycological Institute, Surrey, U.K. 1971.
12. Ellis MB. More Dematiaceous Hyphomycetes. Common wealth Mycological Institute, Surrey, U.K. 1976.
13. Fatimazahra El Bergadi, Faouzi Laachari, Soumya Elabed, Iraqui Houssaini Mohammed & Saad Koraichi Ibensouda. Cellulolytic potential and filter paper activity of fungi isolated from ancient manuscripts from the Medina of Fez. *Ann. Microbiol.* 2013; **64** : 815–822.
14. Garg KL, Kamal K, Mishra AK. Role of fungi in the deterioration of wall paintings. *Sci. Total Environ.* 1995; **167**: 255–271.
15. Gillman JC. A manual of Soil Fungi. Oxford & IBH Publishing Co., Calcutta, 1957.
16. Irene Arroyo. The role of fungi in the deterioration of movable and immovable cultural heritage. *E-Conservation Magazine*, Spain, 2007; 40-50.
17. Kendrick B. Taxonomy of Fungi Imperfecti. University of Toronto Press, Toronto, Canada, 1971.
18. Mesquita N, Portugal A, Videira S, Rodry'guez-Echeverry' S, Bandeira AML, Santos MJA, Freitas H. Fungal diversity in ancient documents. A case study on the archive of the University of Coimbra. *Int. Biodeter. Biodegr.* 2009; **63**: 626–629.
19. Michaelsen A, Piñar G, Montanari M, Pinzari F. Biodeterioration and restoration of a 16th-century book using a combination of conventional and molecular techniques: A case study. *Int. Biodeter. Biodegr.* 2009; **63** : 161–168.
20. Montella S. *et al.* Discovery of genes coding for carbohydrate-active enzyme by metagenomic analysis of lignocellulosic biomasses. *Sci. Rep.* 2017; **7** : 42623.
21. Pinzari F, Pasquariello G, De Mico A. Biodeterioration of paper: a SEM study of fungal spoilage reproduced under controlled conditions. *Macromol. Symp.* 2006; **238** : 57–66.
22. Priyanka P, Yuvraj C, Farha S, Aranganathan V. Isolation of cellulose degrading fungi from soil and optimization for cellulase production using carboxy methyl cellulose. *Int. J. Live Sci. Pharma Res.* 2017; **7** : 57–60.
23. Raper KB, Fennell DI. The Genus *Aspergillus*. The Williams and Wilkins Co., Baltimore, 1965.
24. Raper KB, Thom C, Fennell D. Manual of *Penicillia*. Hafner Publishing Co., New York, 1968.
25. Rifai MA. A Revision of the Genus *Trichoderma*. Mycological Papers No. 116, 1969.

26. Sequeira S, Phillips A, Cabrita E, Macedo M. Antifungal treatment of paper with calcium propionate and parabens: Short-term and long-term effects. *Int. Biodeterior. Biodegradation*. 2017; **120** : 203–215.
27. Srivastava Mamta, Srivastava Neeraj. Fungal deterioration of cultural commodities in paper. *J. Liv. World*. 2007; **14**(1) : 51-53.
28. Srivastava Mamta, Arya MK, Srivastava Neeraj. Cellulolytic fungi causing biodeterioration of Webster's dictionary in Gorakhpur. *International Journal of Biological Technology*. 2011; **2** : 216-220.
29. Sterflinger K, Pinzari F. The revenge of time: fungal deterioration of cultural heritage with particular reference to books, paper and parchment. *Environ. Microbiol*. 2012; **14** : 559–566.
30. Sterflinger K, Piñar G. Microbial deterioration of cultural heritage and works of art — tilting at windmills. *Appl. Microbiol. Biotechnol*. 2013; **97** : 9637–9646.
31. Subramanian CV. Hyphomycetes: Taxonomy and Biology. Academic Press, London, 1983.